DATA REPORT

Water Quality and Stream Invertebrate Assessment

for the Millstone River, Nanaimo, BC,

(Fall 2011)

Report prepared by:

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1. Background

This report documents a water quality and stream invertebrate assessment conducted on the Millstone River, BC, during October-November 2011.

This study was undertaken by undergraduate students attending the Environmental Monitoring (RMOT 306) course at Vancouver Island University (VIU), offered as part of the Bachelor of Natural Resources Protection (Dan McNeill, Clayton Smith, Chris White). Students worked under the supervision of the course instructor, Dr. Eric Demers (Vancouver Island University). This report was compiled by Dr. Eric Demers based on student group reports.

VIU students contributed approximately 125 student-hours to this project, including site visits, project proposal, field sampling, laboratory analyses, and oral and written presentations. Dr. Eric Demers contributed approximately 15 hours for project management and report compilation.

Logistical support was provided by Fisheries and Oceans Canada (DFO). Funding for field expenses and analytical processing of water samples was provided by the Regional District of Nanaimo and Fisheries and Oceans Canada. ALS Laboratory (Burnaby, BC) provided reduced rates on its analytical services for this project.

2. Introduction

The Millstone River watershed encompasses an area of approximately 93 km^2 and is comprised of 26 km of streams, 16 tributaries and 8 lakes. The primary drainage network in the watershed consists of Benson Creek, Brannen Lake and the Millstone River. Benson Creek originates west of Mount Benson at Lucid Lake (elevation: 619 m), and flows in a northerly direction for approximately 12 km into Brannen Lake (elevation: 78 m). The Millstone River flows from Brannen Lake in a southeasterly direction for approximately 14 km to the Strait of Georgia at the Nanaimo Harbour.

During summer 2007, an 800-metre long bypass channel was constructed along the lower Millstone River in Bowen Park by the Nanaimo Fish and Game Protective Association, in partnership with Fisheries and Oceans Canada (DFO) and the City of Nanaimo. The bypass channel was built to provide anadromous salmonids with new spawning and juvenile rearing habitat, and access to the watershed above the Deadman Falls barrier in Bowen Park. Additional habitat restorations were conducted by DFO during 2008 and 2009 to provide summer flow augmentation to the Millstone River and bypass channel.

During summer 2011, gravel was added to the Millstone River to provide returning Coho salmon with more spawning habitat near the Pryde Vista Golf Course.

A need was identified for continued monitoring of water and habitat quality of the Millstone River and bypass channel. Therefore, this report documents a water quality and stream invertebrate assessment conducted on the Millstone River during October-November 2011.

Specific objectives for this study of the Millstone River included:

- establish 6 water quality sampling stations;
- obtain field measurements of water quality at the 6 sampling stations during two sampling events (29 October and 20 November 2011);
- obtain water samples from each sampling station during two sampling events (29 October and 20 November 2011) for detailed laboratory analyses; and,
- collect stream invertebrate samples at 3 sampling stations during one sampling event (29 October 2011) for analysis at Vancouver Island University.

3. Methods

3.1. Study Site

This project was conducted on Benson Creek and the Millstone River located in the City of Nanaimo, BC (Figure 1).

3.1.1. Sampling Stations

Six sampling stations were established on Benson Creek and the Millstone River, during October-November 2011 (Tables 1 and 2; Figure 1). The location of each station was chosen to provide adequate coverage for the length of the Millstone River and to repeat sampling at some of the stations previously used by VIU. Stations were numbered from upstream to downstream (Station 1 on Benson Creek; Stations 2-6 on the Millstone River). All stations were easily accessed via foot paths or road crossings. Station 1 was located on Benson Creek at a crossing on Biggs Road. Stations 2 and 3 were located on the Millstone River at crossings on Biggs Road, and Durnin Road, respectively. Station 4 was located at the end of Pryde Avenue, next to the Pryde Vista Golf Course parking area. Station 5 was located along the bypass channel in Bowen Park, just downstream of the Duck Pond. Station 6 was located in Barsby Park, approximately 175 m upstream of the Millstone estuary. All stations consisted of shallow and gentle riffle sections.

3.1.2. Sampling Schedule

Field sampling was conducted on 29 October and 20 November 2011. For this study, samples were collected for water quality analyses, microbiology and stream invertebrate assessment. Table 2 lists the specific activities conducted at each station during each sampling event. Microbiology and stream invertebrate assessments were only completed during the October sampling event. Photographs showing site conditions are included in Appendix 1.

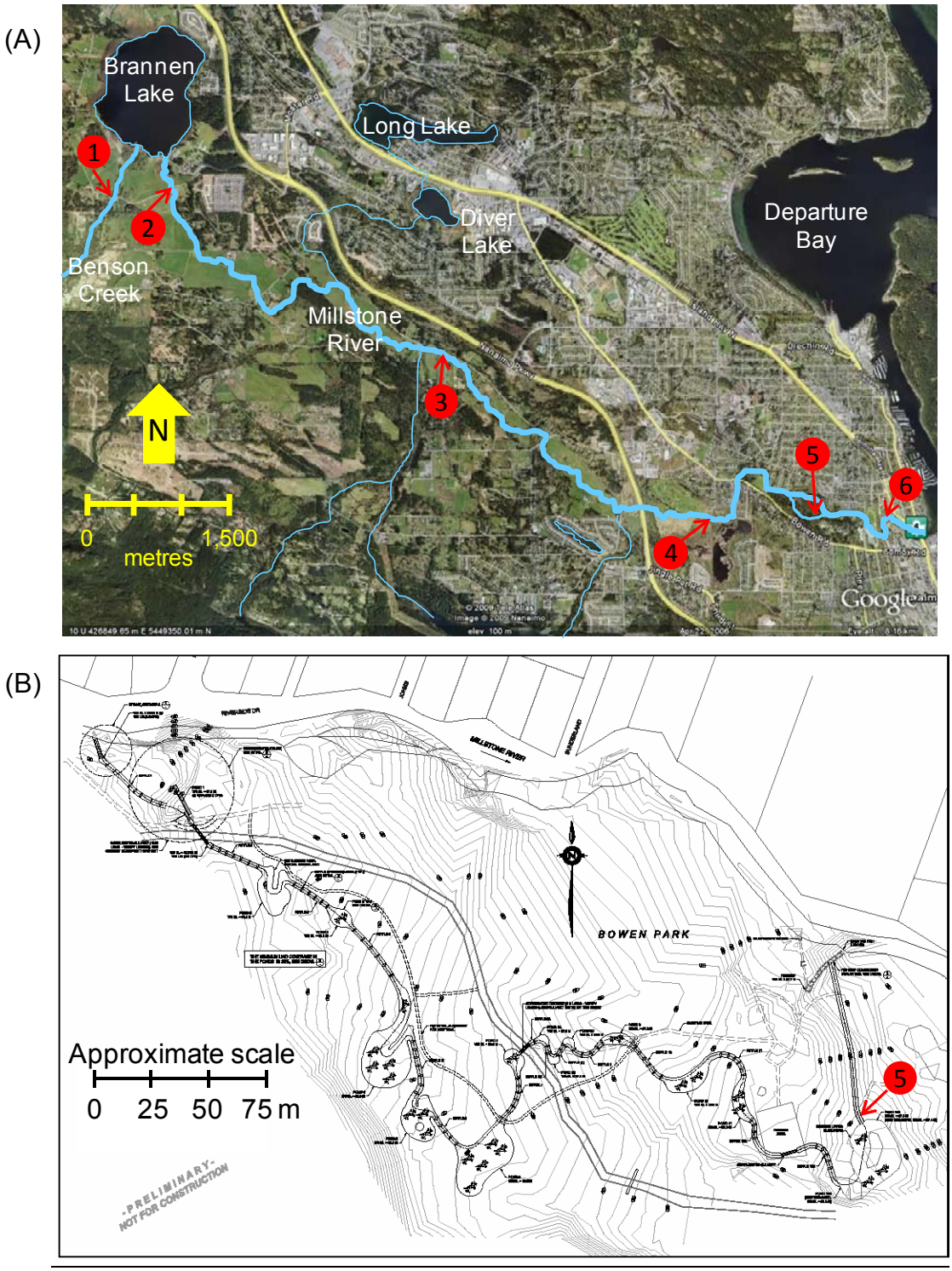

Figure 1. (A) Approximate location of the sampling stations used for water quality and stream invertebrate assessments on the Millstone River during October-November 2011. Station 1 was located on Benson Creek. This map was obtained from Google Earth. (B) Approximate location of sampling stations 5 along the Millstone River Bypass Channel. This map was obtained from Fisheries and Oceans Canada. Table 1 provides details of the specific location of each station. Table 2 details the sampling activities conducted at each station. Map scales are approximated.

Table 1. Description of the sampling stations used for water quality and stream invertebrate assessments on the Millstone River during October-November 2011. Station 1 was located on Benson Creek. All northing and easting coordinates are based on zone 10U and were approximated with Google Earth.

Station	UTM Coordinates		General Location
	Easting	Northing	
1	422738	5450707	Benson Creek, Biggs Road crossing
2	423341	5450828	Millstone River, Biggs Road crossing
3	426304	5448953	Durnin Road crossing
4	429076	5447227	Near Pryde Vista Golf Course parking area
5	430233	5447304	Bypass channel, downstream of Duck Pond
6	430941	5447091	Barsby Park, 170 m upstream from estuary

3.2. Water Quality

3.2.1. Field Measurements

Water quality sampling events were conducted on 29 October and 20 November 2011. At each sampling station, field measurements of water temperature (to the nearest 0.1 ^{o}C), dissolved oxygen (to the nearest 0.01 mg/L) were obtained with a YSI Pro20 electronic probe. The electronic probe was placed directly in the channel water.

Table 2. Water quality and stream invertebrate sampling activities conducted at each station on the Millstone River during October-November 2011. The symbols "A" or "B" indicate whether samples / measurements were taken during the October or November sampling events, respectively.

Station	Water Quality				Stream Invertebrates
	Field Measurements	VIU Analyses	ALS Lab Analyses	Microbiology	
1	A[1], B	A, B	A, B	A	A
2	A[1], B[1]	A, B	A, B	A	A
3	A[1], B	A, B	---	A	---
4	A[1], B	A, B	---	A	---
5	A[1], B[1]	A, B	A, B	A	A
6	A[1], B	A, B	---	A	---

Note: [1] Basic hydrological measurements were collected at all stations during the October sampling event, and at stations 2 and 5 during the November sampling event.

Basic hydrological measurements were taken at stations at all stations on 29 October 2011 and at stations 2 and 5 during 20 November 2011. Water velocity (in m/s) was measured along a 10-m

stream length. An orange was dropped slightly upstream of the stream length and allowed to float downstream through the stream length. A stopwatch was used to measure the travel time of the ball between the upstream and downstream ends of the stream length. The average travel time from 3 passes was used to calculate average water velocity.

Stream wetted widths were measured with a metered tape to the nearest 0.1 m, and wetted depths were measured (along the same wetted widths) with a meter stick to the nearest 0.01 m. Total cross-sectional areas (in m^2) were calculated as the sum of the areas of cross-section polygons. Stream discharge (in m^3/s) was obtained as the product of mean water column velocity and cross sectional area.

3.2.2. Water Sampling

During each sampling event, two sets of water samples were collected for laboratory analyses: one set was transported for analysis at Vancouver Island University (VIU), and another set was shipped for analysis by ALS Laboratory, in Vancouver, BC.

Water samples for analysis at VIU were collected from all stations (Table 2). At each station, a clean pre-labelled 500-ml plastic bottle was rinsed 3 times and then used to collect a water sample (Table 3). All water samples were obtained while standing on the stream bank or within the stream channel by immersing the containers just below the water surface while facing upstream. Care was taken not to disturb the bottom sediments. All water samples were kept in a cooler and stored at approximately 4^oC. Laboratory analyses were conducted at VIU within 48 hours of sampling.

Table 3. Sampling containers and preservatives used for water quality samples taken from the Millstone River during October-November 2011. All containers and preservatives for analysis by ALS Laboratory were provided by ALS Laboratory, Burnaby, BC.

Analytical Parameters	Container	Preservative	Analysed by
Total alkalinity, turbidity	500 ml plastic	None	VIU
Conductivity, pH, total hardness	1 L plastic	None	ALS Laboratory
Nutrients	250 ml amber glass	Sulphuric acid	ALS Laboratory
Total metals	250 ml plastic	Nitric acid	ALS Laboratory

Samples for analysis by ALS Laboratory were collected from stations 1, 2 and 5 during both sampling events (Table 2). At each station, water samples were collected in three clean laboratory-supplied and pre-labelled sample containers (Table 3). All samples were obtained while standing on the stream bank or within the stream channel by directly immersing the containers just below the water surface while facing upstream. Care was taken not to disturb the bottom sediments. Samples for analysis of nutrients and total metals were preserved with laboratory-supplied sulphuric acid and nitric acid, respectively. Bottles with preservatives were

inversed five times for adequate mixing. All water samples were stored in a cooler on site, and shipped with ice packs within 72 hours for laboratory analyses at ALS Laboratory.

3.2.3. VIU Laboratory Analyses

Water samples transported to Vancouver Island University were analysed for conductivity, pH, total alkalinity and turbidity. Conductivity (to the nearest 1 µSiemens/cm) and pH (to the nearest 0.01 pH unit) were obtained with a YSI 556 MPS electronic probe. Total alkalinity (as $CaCO_3$) was measured to the nearest 0.1 mg/L using the HACH AL-DT digital titration method. Turbidity was measured to the nearest 0.01 NTU (Nephelometric Turbidity Units) using a HACH 2100 Potable Turbidimeter.

3.2.4. ALS Laboratory Analyses

Water samples submitted for external analyses were processed as per ALS Laboratory standard analytical procedures. The analytes were: conductivity, total hardness, pH, nutrients (ammonia, nitrite, nitrate, orthophosphate and total phosphorus), and total metals (31 metals).

3.2.5. Quality Assurance / Quality Control

Throughout this study, measures were taken to ensure that potential contamination of water samples was minimized. This included using only clean and rinsed containers, preserving samples as prescribed by the analytical laboratory, and storing collected samples in well-labelled containers.

3.2.6. Data Analyses – Comparison with Applicable Guidelines

Water quality results were compared with the applicable provincial water quality guidelines for the protection of freshwater life. The BC Water Quality Guidelines are the maximum allowable concentration (for potential acute effects) and the 30-day average concentration (for potential chronic effects). All guidelines were obtained from the BC Ministry of Environment, Water Protection Division (http://www.env.gov.bc.ca/wat/wq/).

It is important to note that for some metal parameters, analytical detection limits were above applicable guidelines. These include aluminium, antimony, arsenic, cadmium, chromium, cobalt, copper, lead, nickel, selenium, silver, thallium and vanadium. For these metals, measured values reported to be below method detection limits cannot be assumed to be below the applicable guidelines.

3.3. Microbiology

3.3.1. Field Sampling

Water samples for total and fecal coliform enumeration were collected from each sampling station on 29 October 2011 (Table 2). At each station, a sterile pre-labelled 120-ml Whirl-Pak® bag was used to collect a 100-ml water sample by directly immersing the bag by hand just below

the water surface while facing upstream. All samples were stored in a cooler with ice packs and transported within 48 hours to Vancouver Island University for laboratory analysis.

3.3.2. Laboratory Analyses

In the laboratory, water samples were tested for total coliform and fecal coliform (*Escherichia coli* or *E. coli*) using the m-coliBlue24 membrane filtration method (Millipore Corporation). A 25-ml volume of sample water was filtered through a 47-μm membrane filter (marked with 3-mm gridlines) using a vacuum pump. The filtration apparatus was then rinsed with approximately 5 ml of sterile water. Each membrane filter (including the blank) was then transferred to a Petri plate containing an absorbent pad saturated with m-ColiBlue24 broth. All membrane filters were incubated at 37°C for 20 hours (until bacterial colonies were clearly visible).

Upon completion of the incubation period, membrane filters were then examined for bacterial colonies under a dissection microscope (16X magnification). A red or blue colony represents a total coliform "positive" result (Table 4). A blue colony specifically represents an *E. coli* "positive" result. A clear or white colony represents a total coliform negative result.

All colonies present on a membrane filter were counted and expressed as CFU (colony forming units) per 100-ml of sample water.

Table 4. Possible outcomes of the m-coliBlue24 membrane filtration method.

Bacteria Type	Positive Result	Negative Result
Total coliform	Red or blue colony	Clear or white colony No colony
E. coli	Blue colony only	Non-blue colony

3.4. Stream Invertebrates

3.4.1. Sampling Stations

Stream invertebrate samples were collected from stations 1, 2 and 5 on 29 October 2011 (Table 1; Figure 1). The sampling stations were selected based on hydrological characteristics, apparent substrate uniformity, space available for replicate samples, safety and site access. At the time of sampling, all stations consisted of shallow riffles (water depth ~20-25 cm), with water velocity of ~0.30-0.60 m/s, and primarily sand and gravel substrate.

3.4.2. Invertebrate Sampling

At each station, three replicate samples (triplicates) were obtained using a Hess sampler as per the Pacific Streamkeepers procedures (Taccogna and Munro 1995). Each site was approached by walking from downstream. The invertebrate sampler was hand-pressed into the substrate to

isolate a 0.09-m^2 sampling area. All stones and debris 5 cm or larger within the sampling area were held under water in front of the collecting net and rubbed gently by hand to dislodge invertebrates. Cleaned stones and debris were then placed downstream of the sampling area. The streambed was then gently agitated to a depth of 5 cm to loosen any remaining invertebrates. The content of the collecting net was then transferred into a 125-ml plastic sample jar. The net was carefully inspected to ensure all contents were transferred into the sample jar. Samples were stored in a cooler and transported to Vancouver Island University, where laboratory analyses were completed within 48 hours of sampling.

3.4.3. VIU Laboratory Analyses

Laboratory procedures and identification also followed the Pacific Streamkeepers procedures (Taccogna and Munro 1995). The triplicate samples from each station were combined into a single composite sample per station. The contents of all invertebrate sample jars from a station were poured into a shallow white tray. Invertebrates were sorted into apparent taxonomic groups. Identification to the appropriate taxonomic level (as prescribed by the Pacific Streamkeepers procedures) was confirmed using a dissecting microscope. The number of invertebrates and the number of distinguishable subgroups within each broad taxonomic group were recorded on a Pacific Streamkeeper Invertebrate Survey Field Data Sheet. From these records, various useful metrics were calculated for each station, including: total density (number per m^2), total number of taxonomic groups, predominant taxonomic group, Pollution Tolerance Index, EPT (Ephemeroptera-Plecoptera-Trichoptera) Index, EPT to Total Ratio Index, Predominant Taxon Ratio Index, and overall Site Assessment Rating.

4. Results

Discharge measurements for the Millstone River suggest that water levels were not at bankfull at the time of sampling in October and November 2011 (Table 5). Discharge in Benson Creek was higher than in the Millstone River at Biggs Rd suggesting that the storage capacity of Brannen Lake (source of the Millstone River) had not yet been fully exceeded. The decrease in discharge between stations 3 and 4, and the increase in discharge between stations 4 and 6 may be due to imprecision or error in one of more measurements.

Average air temperature during the 10-day period prior to each sampling event was 7.5^{o}C and 2.7 ^{o}C for the October and November sampling events, respectively (data for Nanaimo Airport retrieved from http://climate.weatheroffice.gc.ca). Total rainfall during the 10-day period prior to the October and November sampling events were 27 mm and 31 mm, respectively.

4.1. Water Quality

4.1.1. Field Measurements and VIU Laboratory Analyses

Water temperature averaged 7.9^{o}C and 4.1^{o}C during the October and November sampling events, respectively (Table 5). The drop in water temperature between sampling events was related to the concurrent decline in air temperature. During both sampling events, the coolest water

occurred in Benson Creek (station 1), and the warmest water occurred in the upper Millstone River (station 2) just downstream from Brannen Lake.

During both sampling events, all dissolved oxygen levels were above the minimum guideline of 9.0 mg/L for early fish life stages (RISC 1998). Overall, dissolved oxygen concentrations were at 88-112% saturation.

Table 5. Field measurements and laboratory results (VIU Laboratory) for water samples taken from six stations on the Millstone River during October-November 2011.

		Field Measurements				VIU Laboratory	
Station	**Discharge (m^3/s)**	**Temperature (^oC)**	**Dissolved Oxygen (mg/L)**	**Conductivity (µS/cm)**	**pH**	**Total Alkalinity (mg/L $CaCO_3$)**	**Turbidity (FAU)**
			29 October 2011				
1	0.73	6.2	13.23	28	7.10	12.4	0.15
2	0.57	10.2	10.21	77	6.70	16.8	0.81
3	0.71	7.9	11.53	117	6.78	32.0	1.45
4	0.41	7.5	11.59	121	6.71	44.0	1.12
5	0.42	7.6	12.79	168	6.45	44.0	1.95
6	2.19	7.7	13.20	162	6.74	50.0	1.61
			20 November 2011				
1		1.8	14.86	14	7.69	10.8	0.24
2	1.48	6.8	10.69	43	7.34	24.0	1.23
3		4.1	13.42	37	7.24	27.6	1.74
4		3.9	13.56	81	7.08	35.6	1.58
5	0.45	3.9	14.33	61	7.06	38.0	1.50
6		4.0	14.49	104	6.98	38.4	1.24

Conductivity averaged 112 and 57 µS/cm during the October and November sampling events, respectively, and increased as expected from upstream to downstream (Table 5). Conductivity decreased by an average of 67% within station between sampling events. This pattern was

similar to observations in previous years (VIU 2009, 2011), and was likely due a dilution effect from increased discharge.

Water pH ranged from 6.45 to 7.69 during this study, and generally decreased from upstream to downstream (Table 5). There was an average increase of 0.49 pH units between sampling events.

Total alkalinity ranged from 10.8 to 50.0 mg/L, and generally decreased between the October and November sampling events at all stations except station 2 (Table 5). There was an increasing trend in alkalinity from upstream to downstream during both sampling events. Total alkalinity in Benson Creek and the Millstone River indicated "moderate acid sensitivity" (10-20 mg/L) and "low acid sensitivity" (≥20 mg/L), respectively, as defined by RISC (1998).

Turbidity levels were below 2 NTU during both sampling events and there was a general increase from upstream to downstream (Table 5).

4.1.2. ALS Laboratory Analyses

Water quality results from ALS Laboratories were compared to the BC Provincial water quality guidelines for the protection of aquatic life (Table 6).

The conductivity measurements from ALS Laboratories were consistent with the field measurements obtained with the electronic probe during the October sampling event, but were lower by an average of 75% during the second sampling event. This discrepancy may have been caused by improper probe calibration. Both sets of conductivity results displayed consistent trends where conductivity increased from upstream to downstream stations, and decreased between sampling events.

Total hardness ranged from 15.0 to 52.2 mg/L, and displayed similar trends as conductivity and total alkalinity (i.e., increase from upstream to downstream, decrease between sampling events). Total hardness was below 60 mg/L during both sampling events, indicating "soft water" as defined by RISC (1998).

The pH measurements from ALS Laboratories span a narrower range of values (7.45-8.03) than the field measurements obtained with the electronic probe. This discrepancies possibly reflects improper probe calibration, differences in air space content among sampling containers and/or time elapsed between sampling and laboratory analysis. All pH levels were within or near the recommended range of 6.5-9.0 for aquatic life (RISC 1998).

Table 6. Laboratory results (ALS Laboratory) for water samples taken from 3 stations on the Millstone River during 29 October and 20 November 2011. All values are expressed in mg/L unless specified otherwise. The values enclosed in boxes exceeded at least one of the applicable water quality guidelines. See additional notes on the next page.

	BC Water Quality Guidelines [a]							
	BC Max	BC 30-day Mean	29 October 2011			20 November 2011		
Variable	mg/L	mg/L	1	2	5	1	2	5
General/Physical								
Conductivity (µS/cm)			48.1	75.0	166	38.7	72.3	144
Hardness, Total			18.8	28.6	52.2	15.0	27.8	43.8
pH (pH units)	6.5 - 9.0		7.89	7.89	8.03	7.45	7.49	7.80
Nutrients								
Ammonia-N	4.64 [b]	0.893 [b]	<0.0050	0.0070	0.0059	<0.0050	0.0109	0.0063
Nitrate (as N)	31.3	3	0.1510	<0.0050	0.0895	0.2230	0.0432	0.1690
Nitrite (as N)	0.06 [c]	0.02 [c]	0.0048	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010
Ortho Phosphate (as P)			<0.0010	<0.0010	0.0015	<0.0010	<0.0010	<0.0010
Total Phosphorus			0.0029	0.0082	0.0138	<0.0020	0.0066	0.0087
Total Metals								
Aluminum (Al) [m]	0.10 [d]	0.05 [d]	<0.20	<0.20	<0.20	<0.20	0.27	<0.20
Antimony (Sb) [m]	0.02		<0.20	<0.20	<0.20	<0.20	<0.20	<0.20
Arsenic (As) [m]	0.005		<0.20	<0.20	<0.20	<0.20	<0.20	<0.20
Barium (Ba)	5	1	<0.010	<0.010	0.018	<0.010	<0.010	0.014
Berylium (Be)	0.0053		<0.0050	<0.0050	<0.0050	<0.0050	<0.0050	<0.0050
Bismuth (Bi)			<0.20	<0.20	<0.20	<0.20	<0.20	<0.20
Boron (B)	1.2		<0.10	<0.10	<0.10	<0.10	<0.10	<0.10
Cadmium (Cd) [m]	0.00001 [e]		<0.010	<0.010	<0.010	<0.010	<0.010	<0.010
Calcium (Ca)			5.26	7.69	15.1	4.18	7.49	12.5
Chromium (Cr) [m]	0.001 [f]		<0.010	<0.010	<0.010	<0.010	<0.010	<0.010
Cobalt (Co) [m]	0.11	0.004	<0.010	<0.010	<0.010	<0.010	<0.010	<0.010
Copper (Cu) [m]	0.003 [g]	0.002 [g]	<0.010	<0.010	<0.010	<0.010	<0.010	<0.010
Iron (Fe)	1.0		<0.030	0.208	0.384	<0.030	0.669	0.236
Lead (Pb) [m]	0.007 [h]	0.004 [h]	<0.050	<0.050	<0.050	<0.050	<0.050	<0.050
Lithium (Li)	0.87	0.096	<0.010	<0.010	<0.010	<0.010	<0.010	<0.010
Magnesium (Mg)			1.37	2.29	3.49	1.11	2.22	3.08
Manganese (Mn)	0.71 [i]	0.67 [i]	<0.0050	0.024	0.017	<0.0050	0.106	0.011
Molybdenum (Mo)	2	1	<0.030	<0.030	<0.030	<0.030	<0.030	<0.030
Nickel (Ni) [m]	0.025 [j]		<0.050	<0.050	<0.050	<0.050	<0.050	<0.050
Phosphorus (P)			<0.30	<0.30	<0.30	<0.30	<0.30	<0.30
Potassium (K)	373		<2.0	<2.0	<2.0	<2.0	<2.0	<2.0
Selenium (Se) [m]		0.002	<0.20	<0.20	<0.20	<0.20	<0.20	<0.20
Silicon (Si)			3.71	2.57	3.25	3.52	3.56	3.86
Silver (Ag) [m]	0.0001 [k]	0.00005 [k]	<0.010	<0.010	<0.010	<0.010	<0.010	<0.010
Sodium (Na)			2.1	3.4	12.2	<2.0	3.4	12.6
Strontium (Sr)			0.020	0.029	0.125	0.016	0.030	0.084
Thallium (Tl) [m]	0.0003		<0.20	<0.20	<0.20	<0.20	<0.20	<0.20
Tin (Sn)			<0.030	<0.030	<0.030	<0.030	<0.030	<0.030
Titanium (Ti)	2		<0.010	<0.010	<0.010	<0.010	0.015	<0.010
Vanadium (V) [m]	0.006		<0.030	<0.030	<0.030	<0.030	<0.030	<0.030
Zinc (Zn)	0.033 [l]	0.0075 [l]	<0.0050	<0.0050	<0.0050	<0.0050	<0.0050	<0.0050

Table 6. (Continued)

NOTES:

Results are expressed as mg/L except for pH and conductivity.
"<" means less than the detection limit.

[a] BC Water Quality Guidelines (WQG) compiled from
http://www.env.gov.bc.ca/wat/wq/wq_guidelines.html
http://www.env.gov.bc.ca/wat/wq/BCguidelines/working.html

[b] Total ammonia guideline is dependent on water temperature and pH of tested water.

[c] Nitrite guideline is for chloride concentration < 2 mg/L.

[d] Aluminum guidelines for pH ≥ 6.5.

[e] The maximum cadmium guideline is $0.001 * 10^{\{0.86 [\log(\text{hardness})] - 3.2\}}$ mg/L.

[f] Chromium guideline is for the more toxic Chromium VI.

[g] The maximum copper guideline is 0.001 * [0.094(hardness) + 2] mg/L.
The 30-day mean copper guideline is for hardness < 50 mg/L.

[h] The maximum lead guideline is $0.001 * e^{\{1.273 [\ln(\text{hardness})] - 1.46\}}$ mg/L.
The 30-day mean lead guideline is $0.001 * [3.31 + e^{\{1.273 [\ln(\text{hardness})] - 4.704\}}]$ mg/L.

[i] The maximum manganese guideline is 0.01102 * (hardness) + 0.54 mg/L.
The 30-day mean manganese guideline is 0.0044 * (hardness) + 0.605 mg/L.

[j] Nickel guideline is for hardness < 60 mg/L.

[k] Silver guidelines are for hardness < 100 mg/L.

[l] Zinc guidelines are for hardness < 90 mg/L.

[m] Analytical detection limits were above applicable guidelines for these metals.

All nutrient levels were below applicable guidelines and/or below detection limits. Total ammonia was below detection limit (i.e., <0.005 mg/L) in Benson Creek, and ranged from 0.0059 to 0.0109 mg/L in the Millstone River. Nitrate concentrations ranged from below detection limit to 0.223 mg/L during this study. During both sampling events, the highest nitrate levels occurred at station 1 (Benson Creek). Similar high nitrate levels have been observed repeatedly at this location during other water quality monitoring projects (VIU 2009, 2010; E. Demers, unpublished data; 2004-2010). All nitrite levels were below detection limit, except in Benson Creek during the October sampling event (i.e., 0.0048 mg/L).

Orthophosphate was below or near detection limit (i.e., <0.0015 mg/L) during both sampling events. Total phosphorus ranged from below or near detection limit to 0.0138 mg/L during this study. There was an increase from upstream to downstream stations, and a decreased between sampling events. Overall, total phosphorus levels were mainly within or near the low range of <0.010 mg/L typical of "oligotrophic" waters as defined by RISC (1998).

All metal concentrations were below the applicable water quality guidelines and/or below detection limits, except aluminium at station 2 during the November sampling event. Total metal analyses measure the combined amount of metals dissolved in water and bound to particles. In general, dissolved metals are more bio-available (hence toxicologically available) than metals that are bound to particles. It is unclear whether the observed elevated aluminium represented dissolved metals or metals bound to suspended particles.

4.2. Microbiology

All samples collected from the Millstone River contained some coliform bacteria (Table 7). Total coliform and *E. coli* levels generally increased from upstream to downstream, with the highest levels observed at station 4 (total coliform: 516 CFU / 100 ml; *E. coli*: 356 CFU / 100 ml). The proportion of total coliform made up of *E. coli* bacteria also generally increased from upstream to downstream.

Table 7. Total coliform and *E. coli* counts from water samples taken at six stations on the Millstone River during 29 October 2011. All values are expressed as CFU (colony forming units) per 100 ml. No microbiology samples were collected during 20 November 2011.

Station	Total Coliform	*E. coli*	% *E. coli*
1	112	0	0.0%
2	252	64	25.4%
3	224	116	51.8%
4	516	356	69.0%
5	396	260	65.7%
6	312	204	65.4%

4.3. Stream Invertebrates

A total of 698 stream invertebrates representing 9 broad taxonomic groups were counted at three stations on the Millstone River during 29 October 2011 (Table 8; Figure 2; Appendix 2). Animal density increased from upstream to downstream stations, with the highest density observed at station 5 located along the bypass channel where density reached 1,533 animals/m^2. Caddisfly larvae, mayfly nymphs and aquatic worms (oligochaetes) were the most common taxonomic group, although their abundance was variable among stations.

Site assessment ratings ranged from 2.25 to 3.25 suggesting "marginal" (station 1) to "acceptable" (stations 2 and 5) invertebrate community abundance and diversity. The representation of pollution-sensitive mayfly nymphs, stonefly nymphs and caddisfly larvae (EPT taxa) at station 5 suggests continued adequate invertebrate colonization in the bypass channel.

Table 8. Abundance and density of stream invertebrates obtained from triplicate samples taken at three stations on the Millstone River during 29 October 2011. Overall site assessment ratings are also provided for each station (out of a possible range of 1.00-4.00). Invertebrate Survey Field Data Sheets are included in Appendix 2. No stream invertebrate samples were collected on 20 November 2011.

Pollution Tolerance	Invertebrate Taxa	Station 1	Station 2	Station 5
Category 1 Pollution Intolerant	Caddisfly Larva	5	127	114
	Mayfly Nymph	0	0	169
	Stonefly Nymph	32	0	7
	Dobsonfly (hellgrammite)	0	1	0
Category 2 Somewhat Pollution Intolerant	Clam, Mussel	0	0	3
	Scud (Amphipod)	11	32	62
Category 3 Pollution Tolerant	Aquatic Worm (oligochaete)	34	42	57
	Leech	0	0	1
	Midge Larva (chironomid)	0	1	0
	Total Abundance	82	203	413
	Density (number / m^2)	304	752	1,533
	Site Assessment Rating	2.75	2.25	3.25

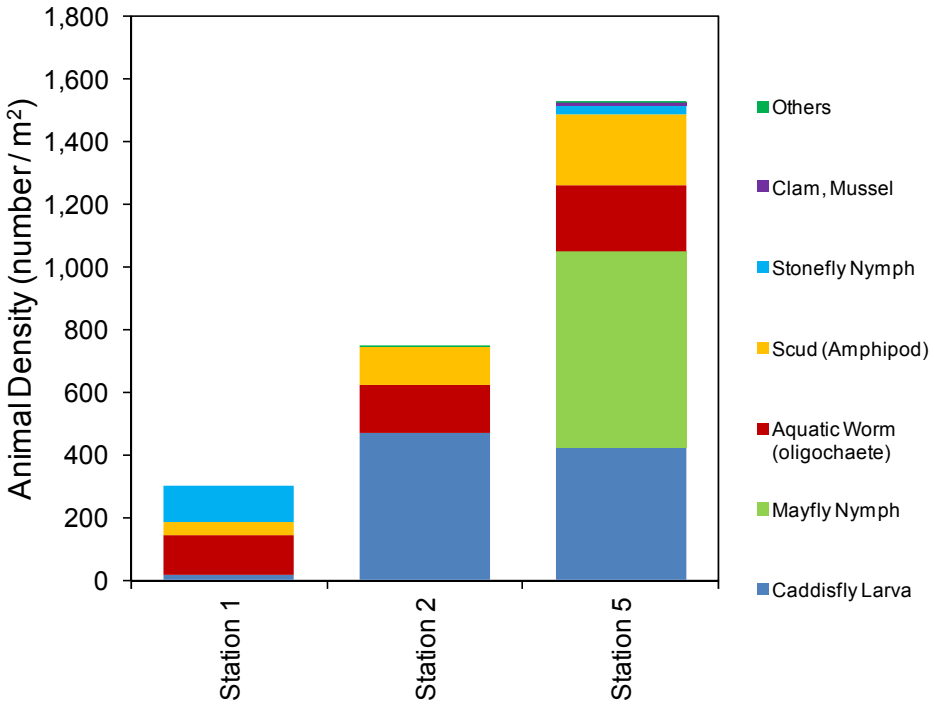

Figure 2. Density of stream invertebrates obtained from triplicate samples taken at three stations on the Millstone River during 29 October 2011. The "Other" category includes dobsonfly larva, leech and midge larva (chironomid). Data are summarized in Table 8 and Invertebrate Survey Field Data Sheets are included in Appendix 2.

5. Acknowledgements

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6. References

RISC. 1998. Guidelines for Interpreting Water Quality Data. Resources Information Standards Committee, Victoria, BC.

Taccogna, G., and K. Munro (eds). 1995. The Streamkeepers Handbook: a Practical Guide to Stream and Wetland Care. Salmonid Enhancement Program, Dept. Fisheries and Oceans, Vancouver, BC.

Vancouver Island University (VIU: B. Brooks, M. Fuller, L. Isaac, J. Patterson, B. Simmons, L. Sobie, and E. Demers). 2009. Water Quality and Stream Invertebrate Assessment for the Millstone River, Nanaimo, BC (Fall 2008). Data Report.

Vancouver Island University (VIU: A. Goeppel, N. Hamilton, A. Kennedy, F. Raffaelli, J. Sellars, S. Vickers, and E. Demers). 2010. Water Quality and Stream Invertebrate Assessment for the Millstone River, Nanaimo, BC (Fall 2009). Data Report.

Vancouver Island University (VIU: D. Clark, A. Hile, C. McCulloch, and E. Demers). 2011. Water Quality and Stream Invertebrate Assessment for the Millstone River, Nanaimo, BC (Fall 2010). Data Report.

7. Appendices

APPENDIX 1. Photographs showing site conditions at each sampling station on the Millstone River on 29 October 2011 (photographs by Chris White).

Photo 1. Upstream view of Benson Creek at the Biggs Road crossing (station 1).

Photo 2. Downstream view of the Millstone River at the Biggs Road crossing (station 2).

APPENDIX 1. (Continued)

Photo 3. Downstream view of the Millstone River at the Durnin Road crossing (station 3).

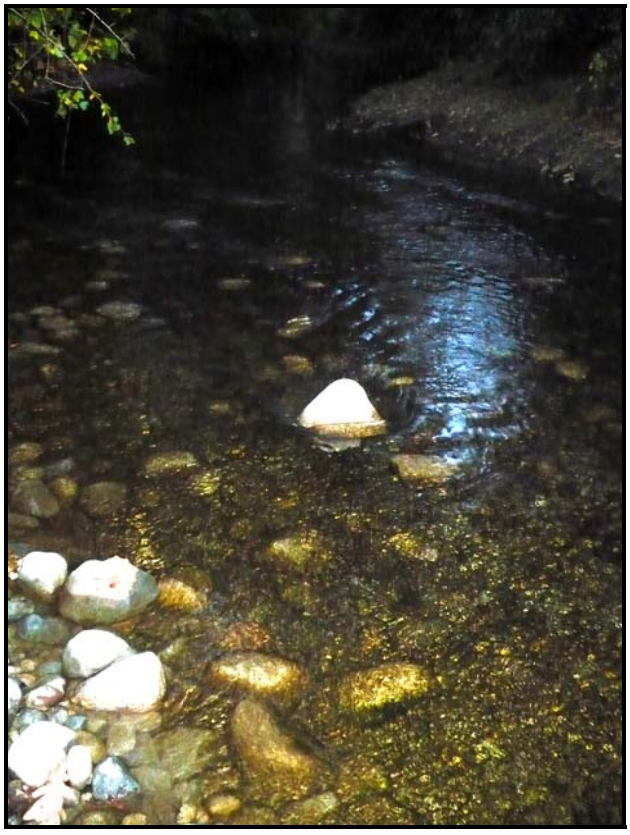

Photo 4. Downstream view of the Millstone River near the Pryde Vista Golf Course (station 4).

APPENDIX 1. (Continued)

Photo 5. Millstone bypass channel at the outlet of the Duck Pond (station 5).

Photo 6. Upstream view of the Millstone River in Barsby Park (station 6).

APPENDIX 2. Invertebrate Survey Field Data Sheet completed for triplicate stream invertebrate samples collected at Stations 1, 2 and 5 on the Millstone River on 29 October 2011.

INVERTEBRATE SURVEY FIELD DATA SHEET (Page 1 of 2)

Stream Name:	Benson Creek		Date: 29 October 2011
Station Name:	**Station 1**		Flow status: Moderate
Sampler Used: Hess	Number of replicates 3	Total area sampled (Hess, Surber = 0.09 m^2) x no. replicates 0.09 x 3 = 0.27 m^2	

Column A Pollution Tolerance	Column B Common Name	Column C Number Counted	Column D Number of Taxa
Category 1 **Pollution Intolerant**	Caddisfly Larva (EPT)	5	2
	Mayfly Nymph (EPT)		
	Stonefly Nymph (EPT)	32	3
	Dobsonfly (hellgrammite)		
	Gilled Snail		
	Riffle Beetle		
	Water Penny		
Sub-Total		37	5
Category 2 **Somewhat Pollution Tolerant**	Alderfly Larva		
	Aquatic Beetle		
	Aquatic Sowbug		
	Clam, Mussel		
	Cranefly Larva		
	Crayfish		
	Damselfly Larva		
	Dragonfly Larva		
	Fishfly Larva		
	Scud (amphipod)	11	1
	Watersnipe Larva		
Sub-Total		11	1
Category 3 **Pollution Tolerant**	Aquatic Worm (oligochaete)	34	2
	Blackfly Larva		
	Leech		
	Midge Larva (chironomid)		
	Planarian (flatworm)		
	Pouch and Pond Snails		
	True Bug Adult		
	Water Mite		
Sub-Total		34	2
TOTAL		82	8

APPENDIX 2. (Continued)

INVERTEBRATE SURVEY INTERPRETATION SHEET (Page 2 of 2)

SECTION 1 - ABUNDANCE AND DENSITY

ABUNDANCE: Total number of organisms from cell **CT**: 82

DENSITY: Invertebrate density per square metre:

82 ÷ 0.27 = 304

PREDOMINANT TAXON:

Invertebrate group with the highest number counted (Col. C): Aquatic Worm (oligochaete)

SECTION 2 - WATER QUALITY ASSESSMENTS

POLLUTION TOLERANCE INDEX: Sub-total number of taxa found in each tolerance category.

Good	Accpetable	Marginal	Poor
>22	17-22	11-16	<11

3 x D1 + 2 x D2 + D3

3 x __5__ + 2 x __1__ + __2__ = 19

EPT INDEX: Total number of EPT taxa.

Good	Accpetable	Marginal	Poor
>8	5-8	2-4	0-1

EPT4 + EPT5 + EPT6

__2__ + __0__ + __3__ = 5

EPT TO TOTAL RATIO INDEX: Total number of EPT organisms divided by the total number of organisms.

Good	Accpetable	Marginal	Poor
0.75-1.0	0.50-0.74	0.25-0.49	<0.25

(EPT1 + EPT2 + EPT3) / CT

(5 + 0 + 32) / 82 = 0.45

SECTION 3 - DIVERSITY

TOTAL NUMBER OF TAXA: Total number of taxa from cell **DT**: 8

PREDOMINANT TAXON RATIO INDEX: Number of invertebrate in the **predominant taxon** (S3) divided by CT.

Good	Accpetable	Marginal	Poor
<0.40	0.40-0.59	0.60-0.79	0.80-1.0

Col. C for S3 / CT

_34_ / _82_ = 0.41

SECTION 4 - OVERALL SITE ASSESSMENT RATING

SITE ASSESSMENT RATING: Assign a rating of 1-4 to each index (S4, S5, S6, S8), then calculate the average.

Assessment Rating	
Good	4
Accpetable	3
Marginal	2
Poor	1

Assessment	Rating
Pollution Tolerance Index	3
EPT Index	3
EPT To Total Ratio	2
Predominant Taxon Ratio	3

Average Rating
2.75

APPENDIX 2. (Continued)

INVERTEBRATE SURVEY FIELD DATA SHEET (Page 1 of 2)

Stream Name:	Millstone River		Date:	29 October 2011
Station Name:	**Station 2**		Flow status:	Moderate
Sampler Used:	Number of replicates	Total area sampled (Hess, Surber = 0.09 m^2) x no. replicates		
Hess	3	0.09 x 3 =	0.27	m^2

Column A Pollution Tolerance	Column B Common Name	Column C Number Counted	Column D Number of Taxa
Category 1 **Pollution Intolerant**	Caddisfly Larva (EPT)	127	2
	Mayfly Nymph (EPT)		
	Stonefly Nymph (EPT)		
	Dobsonfly (hellgrammite)	1	1
	Gilled Snail		
	Riffle Beetle		
	Water Penny		
Sub-Total		128	3
Category 2 **Somewhat Pollution Tolerant**	Alderfly Larva		
	Aquatic Beetle		
	Aquatic Sowbug		
	Clam, Mussel		
	Cranefly Larva		
	Crayfish		
	Damselfly Larva		
	Dragonfly Larva		
	Fishfly Larva		
	Scud (amphipod)	32	1
	Watersnipe Larva		
Sub-Total		32	1
Category 3 **Pollution Tolerant**	Aquatic Worm (oligochaete)	42	2
	Blackfly Larva		
	Leech		
	Midge Larva (chironomid)	1	1
	Planarian (flatworm)		
	Pouch and Pond Snails		
	True Bug Adult		
	Water Mite		
Sub-Total		43	3
TOTAL		203	7

APPENDIX 2. (Continued)

INVERTEBRATE SURVEY INTERPRETATION SHEET (Page 2 of 2)

SECTION 1 - ABUNDANCE AND DENSITY

ABUNDANCE: Total number of organisms from cell **CT**: 203

DENSITY: Invertebrate density per square metre:

203 ÷ 0.27 = 752

PREDOMINANT TAXON:

Invertebrate group with the highest number counted (Col. C): Caddisfly Larva (EPT)

SECTION 2 - WATER QUALITY ASSESSMENTS

POLLUTION TOLERANCE INDEX: Sub-total number of taxa found in each tolerance category.

Good	Accpetable	Marginal	Poor
>22	17-22	11-16	<11

3 x D1 + 2 x D2 + D3

3 x __3__ + 2 x __1__ + __3__ = 14

EPT INDEX: Total number of EPT taxa.

Good	Accpetable	Marginal	Poor
>8	5-8	2-4	0-1

EPT4 + EPT5 + EPT6

__2__ + __0__ + __0__ = 2

EPT TO TOTAL RATIO INDEX: Total number of EPT organisms divided by the total number of organisms.

Good	Accpetable	Marginal	Poor
0.75-1.0	0.50-0.74	0.25-0.49	<0.25

(EPT1 + EPT2 + EPT3) / CT

(127 + 0 + 0) / 203 = 0.63

SECTION 3 - DIVERSITY

TOTAL NUMBER OF TAXA: Total number of taxa from cell **DT**: 7

PREDOMINANT TAXON RATIO INDEX: Number of invertebrate in the **predominant taxon** (S3) divided by CT.

Good	Accpetable	Marginal	Poor
<0.40	0.40-0.59	0.60-0.79	0.80-1.0

Col. C for S3 / CT

_127_ / _203_ = 0.63

SECTION 4 - OVERALL SITE ASSESSMENT RATING

SITE ASSESSMENT RATING: Assign a rating of 1-4 to each index (S4, S5, S6, S8), then calculate the average.

Assessment Rating	
Good	4
Accpetable	3
Marginal	2
Poor	1

Assessment	Rating
Pollution Tolerance Index	2
EPT Index	2
EPT To Total Ratio	3
Predominant Taxon Ratio	2

Average Rating
2.25

APPENDIX 2. (Continued)

INVERTEBRATE SURVEY FIELD DATA SHEET (Page 1 of 2)

Stream Name:	Millstone River		Date:	29 October 2011
Station Name:	**Station 5**		Flow status:	Moderate
Sampler Used:	Number of replicates	Total area sampled (Hess, Surber = 0.09 m^2) x no. replicates		
Hess	3	0.09 x 3 =	0.27	m^2

Column A Pollution Tolerance	Column B Common Name	Column C Number Counted	Column D Number of Taxa
Category 1 Pollution Intolerant	Caddisfly Larva (EPT)	114	2
	Mayfly Nymph (EPT)	169	2
	Stonefly Nymph (EPT)	7	2
	Dobsonfly (hellgrammite)		
	Gilled Snail		
	Riffle Beetle		
	Water Penny		
Sub-Total		290	6
Category 2 Somewhat Pollution Tolerant	Alderfly Larva		
	Aquatic Beetle		
	Aquatic Sowbug		
	Clam, Mussel	3	1
	Cranefly Larva		
	Crayfish		
	Damselfly Larva		
	Dragonfly Larva		
	Fishfly Larva		
	Scud (amphipod)	62	1
	Watersnipe Larva		
Sub-Total		65	2
Category 3 Pollution Tolerant	Aquatic Worm (oligochaete)	57	2
	Blackfly Larva		
	Leech	1	1
	Midge Larva (chironomid)		
	Planarian (flatworm)		
	Pouch and Pond Snails	1	1
	True Bug Adult		
	Water Mite		
Sub-Total		59	4
TOTAL		414	12

APPENDIX 2. (Continued)

INVERTEBRATE SURVEY INTERPRETATION SHEET (Page 2 of 2)

SECTION 1 - ABUNDANCE AND DENSITY

ABUNDANCE: Total number of organisms from cell **CT**: 414

DENSITY: Invertebrate density per square metre:

414 ÷ 0.27 = 1533

PREDOMINANT TAXON:

Invertebrate group with the highest number counted (Col. C): Mayfly Nymph (EPT)

SECTION 2 - WATER QUALITY ASSESSMENTS

POLLUTION TOLERANCE INDEX: Sub-total number of taxa found in each tolerance category.

Good	Accpetable	Marginal	Poor
>22	17-22	11-16	<11

3 x D1 + 2 x D2 + D3

3 x __6__ + 2 x __2__ + __4__ = 26

EPT INDEX: Total number of EPT taxa.

Good	Accpetable	Marginal	Poor
>8	5-8	2-5	0-1

EPT4 + EPT5 + EPT6

__2__ + __2__ + __2__ = 6

EPT TO TOTAL RATIO INDEX: Total number of EPT organisms divided by the total number of organisms.

Good	Accpetable	Marginal	Poor
0.75-1.0	0.50-0.74	0.25-0.49	<0.25

(EPT1 + EPT2 + EPT3) / CT

(114 + 169 + 7) / 414 = 0.70

SECTION 3 - DIVERSITY

TOTAL NUMBER OF TAXA: Total number of taxa from cell **DT**: 12

PREDOMINANT TAXON RATIO INDEX: Number of invertebrate in the **predominant taxon** (S3) divided by CT.

Good	Accpetable	Marginal	Poor
<0.40	0.40-0.59	0.60-0.79	0.80-1.0

Col. C for S3 / CT

_169_ / _414_ = 0.41

SECTION 4 - OVERALL SITE ASSESSMENT RATING

SITE ASSESSMENT RATING: Assign a rating of 1-4 to each index (S4, S5, S6, S8), then calculate the average.

Assessment Rating	
Good	4
Accpetable	3
Marginal	2
Poor	1

Assessment	Rating
Pollution Tolerance Index	4
EPT Index	3
EPT To Total Ratio	3
Predominant Taxon Ratio	3

Average Rating
3.25